

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEEENNE POUR LA PROTECTION DES PLANTES**  
(11-17239)

**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Target Organism</b>	Phyllosticta citricarpa	
<b>Short description</b>	Detection of Phyllosticta citricarpa by PCR (Bonants et al. 2003)	
<b>Laboratory contact details</b>	Council for Agricultural Research and Economics- Research Centre for Plant Protection and Certification Via Carlo Giuseppe Bertero, 22, 00156 Rome, Italy	
<b>Date and reference of the validation report</b>	2014-09-15 -	
<b>Validation process according to EPPO Standard PM 7/98:</b>	Yes	
<b>Reference of the test description</b>	0 Bonants P.J.M., Caroll G.C., de Weerd M., van Brouwershaven I.R., Baayen R.P., 2003. Development and validation of a fast PCR-based method for pathogenic isolates of the citrus black spot fungus Guignardia citricarpa. European Journal of Plant Pathology, 109, 503-513. van Gent-Pelzer M.P.E., van Brouwershaven I.R., Kox L.F.F., Bonants P.J.M., 2007. A Taqman PCR method for routine diagnosis of the quarantine fungus Guignardia citricarpa. Journal of Phytopathology, 155, 357-363. EPPO PM 7/17(2), 2009. Guignardia citricarpa. EPPO Bulletin, 39, 318-327.	
<b>Is the test the same as described in the EPPO DP?</b>	Yes	
<b>Is the lab accredited for this test?</b>	No	
<b>Plant species tested (if relevant)</b>	Citrus lemon	
<b>Matrices tested (if relevant)</b>	Fungal mycelium, fruits	
<b>List of methods used</b>		
<b>Method for extraction / isolation / baiting of target organism from matrix</b>		
<b>Molecular methods, e.g. hybridization, PCR and real time PCR</b>	X	PCR method
<b>Serological methods: IF, ELISA, Direct Tissue Blot Immuno Assay</b>		
<b>Plating methods: selective isolation</b>		

<b>Bioassay methods: selective enrichment in host plants, baiting, plant test and grafting.</b>		
<b>Pathogenicity test</b>		
<b>Fingerprint methods: protein profiling, fatty acid profiling &amp; DNA profiling</b>		
<b>Morphological and morphometrical methods intended for identification</b>		
<b>Biochemical methods: e.g. enzyme electrophoresis, protein profiling</b>		
<b>Other</b>		
<b><u>Analytical sensitivity (= limit of detection)</u></b>		
<b>What is smallest amount of target that can be detected reliably?</b>	20 pg of DNA	
<b><u>Diagnostic sensitivity</u></b>		
<b>Proportion of infected/infested samples tested positive compared to results from the standard test , see appendix 2 of PM 7/98</b>		
<b>Specify the standard test</b>		
<b><u>Analytical specificity</u></b>		
<b>Specificity value</b>		
<b>Number of strains/populations of target organisms tested</b>	4 target strains for PCR	
<b>Number of non-target organisms tested</b>	3 non-target strains (see validation report)	
<b>Cross reacts with (specify the species)</b>	No cross reaction	
<b><u>Diagnostic Specificity</u></b>		
<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>		
<b>Specify the standard test</b>		
<b><u>Reproducibility</u></b>		
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100%	
<b><u>Repeatability</u></b>		
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100%	

<b>Test performance study</b>	
<b>Test performance study?</b>	Yes
<b>Include brief details of the test performance study and its output. If available, provide a link to published article/report</b>	<p>The robustness of the method was verified through a Test Performance Study among 7 laboratories. For each lab 6 positive samples (3 containing the target DNA slightly above the relative limit of detection and 3 containing the target DNA ten times the relative limit of detection) and 6 negative samples (3 containing no DNA and 3 containing DNA of non-target strains) were tested. The results showed:</p> <ul style="list-style-type: none"> <li>-100% relative sensitivity</li> <li>-100% relative specificity</li> <li>-100% repeatability</li> <li>-100% reproducibility</li> </ul>
<b>Other information</b>	
<b>Any other information considered useful e.g. robustness, ease of performing the test, etc.</b>	<p>The verification of performance criteria did not give the same value as the limit of detection defined in the EPPO bulletin PM 7/17 (20 pg instead of 1pg), so a new validation was performed</p> <p>See full report for details or contact lab1</p>
The following complementary files are available online:	<ul style="list-style-type: none"> <li>• <a href="#">Validation process of the conventional PCR for the identification of <i>Phyllosticta citricarpa</i> (Bonants et al., 2003)</a></li> </ul>