

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES**  
(11-17239)

**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Target Organism</b>	Globodera pallida Globodera rostochiensis	
<b>Short description</b>	Diagnosis of Globodera pallida and Globodera rostochiensis (potato cyst nematodes) using Taqman® real-time PCR	
<b>Laboratory contact details</b>	Fera Sand Hutton, YO41 1LZ York, United Kingdom	
<b>Date and reference of the validation report</b>	2013-09 - Potato Council Project Report 2009/15: Validation of quantitative DNA detection systems for PCN. Ref: R287	
<b>Validation process according to EPPO Standard PM 7/98:</b>	Yes	
<b>Reference of the test description</b>	PM 7/040 Based on: Bulman S. R. & Marshall J. W. (1997) Differentiation of Australasian potato cyst nematode (PCN) populations using the polymerase chain reaction (PCR). New Zealand Journal of Crop and Horticultural Science 25: 123-129.	
<b>Is the test the same as described in the EPPO DP?</b>	Yes	
<b>Is the lab accredited for this test?</b>	Yes	
<b>Plant species tested (if relevant)</b>		
<b>Matrices tested (if relevant)</b>	Dissected cyst and eggs. Half is retained if morphological examination is required	
<b>List of methods used</b>		
<b>Method for extraction / isolation / baiting of target organism from matrix</b>	X	Conventional flotation method to isolate cysts (Wye washer, following EPPO diagnostic protocol PM 7/40 (3))
<b>Molecular methods, e.g. hybridization, PCR and real time PCR</b>	X	Spin column based DNA extraction and Real-time PCR PCR developed from Bulman S. R. & Marshall J. W. (1997) Applied Biosystems TaqMan Universal master mix II, no UNG (4440043) 50°C for 2 min; 95°C for 10 min; 95°C 15 sec, 60°C 1 min (40 repeats)
<b>Serological methods: IF, ELISA, Direct Tissue Blot Immuno Assay</b>		
<b>Plating methods: selective isolation</b>		
<b>Bioassay methods: selective</b>		

<b>enrichment in host plants, baiting, plant test and grafting.</b>		
<b>Pathogenicity test</b>		
<b>Fingerprint methods: protein profiling, fatty acid profiling &amp; DNA profiling</b>		
<b>Morphological and morphometrical methods intended for identification</b>		
<b>Biochemical methods: e.g. enzyme electrophoresis, protein profiling</b>		
<b>Other</b>		
<b>Analytical sensitivity (= limit of detection)</b>		
<b>What is smallest amount of target that can be detected reliably?</b>	DNA from single cyst detectable at 1000 fold dilution	
<b>Diagnostic sensitivity</b>		
<b>Proportion of infected/infested samples tested positive compared to results from the standard test , see appendix 2 of PM 7/98</b>	G. pallida 100% G. rostochiensis 100%	
<b>Specify the standard test</b>	The TaqMan assay was compared to the standard conventional PCR assay of Bulman & Marshall, 1997.	
<b>Analytical specificity</b>		
<b>Specificity value</b>		
<b>Number of strains/populations of target organisms tested</b>	20+ strains of G. pallida (see validation report) 20+ strains of G. rostochiensis (see validation report)	
<b>Number of non-target organisms tested</b>	Strains of G. tabacum (see validation report) Strains of G. achillae/millefolii (see validation report)	
<b>Cross reacts with (specify the species)</b>	G. tabacum, however this gives a distinct profile and be differentiated for G.pallida and G.rostochiensis	
<b>Diagnostic Specificity</b>		
<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>	G. pallida 87.1% G. rostochiensis 93.75% The testing gave no false negatives for either species.	
<b>Specify the standard test</b>	The TaqMan assay was compared to the standard conventional PCR assay of Bulman & Marshall, 1997 and morphological identification.	
<b>Reproducibility</b>		
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	The testing has been successful carried out by multiple users on all equipment over several days	
<b>Repeatability</b>		
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	Positive controls used on every run produce repeatable results.	

<b>Test performance study</b>	
<b>Test performance study?</b>	Yes
<b>Include brief details of the test performance study and its output. If available, provide a link to published article/report</b>	100% correct identification results over recent proficiency tests.
<b>Other information</b>	
<b>Any other information considered useful e.g. robustness, ease of performing the test, etc.</b>	The <i>Globodera pallida</i> probe is known to cross-react slightly with <i>Globodera rostochiensis</i> DNA. The cross reaction will show as a slight increase in delta Rn in the FAM channel ( <i>G. pallida</i> ) as the delta Rn increases exponentially in the TET channel ( <i>G. rostochiensis</i> ). This cross reaction is only observed when a sample is positive for <i>G. rostochiensis</i> .
The following complementary files are available online:	<ul style="list-style-type: none"> <li>• <a href="#">Validation of quantitative DNA detection systems for PCN Ref: R287</a></li> <li>• <a href="#">Validation report: speciation of <i>Globodera pallida</i> and <i>G. rostochiensis</i> (potato cyst nematodes) using TaqMan® real-time PCR</a></li> </ul>