

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES**  
(11-17239)

**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

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| Target Organism  | Xanthomonas axonopodis pv. dieffenbachiae   |  |
| Short description  | Detection of Xanthomonas axonopodis pv. dieffenbachiae by nested-PCR in leaves and pure culture   |  |
| Laboratory contact details   | Anses Plant Health Laboratory - Pests and Tropical Pathogens Unit<br>Pôle de Protection des Plantes, 7 Chemin de l'IRAT, 97410 Saint Pierre, France |  |
| Date and reference of the validation report  | 2012-03 - Inter-laboratory ring test : Xanthomonas axonopodis pv. dieffenbachiae in Anthurium (Report Xad01-version 2)                              |  |
| Validation process according to EPPO Standard PM 7/98:                                   | No  |  |
| Reference of the test description  | N/R   |  |
| Is the test the same as described in the EPPO DP?  | Yes   |  |
| Is the lab accredited for this test?   | No  |  |
| Plant species tested (if relevant)   | Anthurium sp.   |  |
| Matrices tested (if relevant)  | Leaves and pure culture   |  |
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| List of methods used   |   |  |
| Method for extraction / isolation / baiting of target organism from matrix               | X   | Extraction as in Appendix 1 of PM7/23(2) |
| Molecular methods, e.g. hybridization, PCR and real time PCR                             | X   | Nested-PCR as in Appendix 4 of PM7/23(2) |
| Serological methods: IF, ELISA, Direct Tissue Blot Immuno Assay                          |   |  |
| Plating methods: selective isolation   |   |  |
| Bioassay methods: selective enrichment in host plants, baiting, plant test and grafting. |   |  |
| Pathogenicity test   |   |  |
| Fingerprint methods: protein profiling, fatty acid profiling & DNA                       |   |  |

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| <b>profiling</b>  |   |  |
| <b>Morphological and morphometrical methods intended for identification</b>   |   |  |
| <b>Biochemical methods: e.g. enzyme electrophoresis, protein profiling</b>  |   |  |
| <b>Other</b>  |   |  |
| <b><u>Analytical sensitivity (= limit of detection)</u></b>   |   |  |
| <b>What is smallest amount of target that can be detected reliably?</b>   | 1x10 <sup>4</sup> CFU.mL-1  |  |
| <b><u>Diagnostic sensitivity</u></b>  |   |  |
| <b>Proportion of infected/infested samples tested positive compared to results from the standard test , see appendix 2 of PM 7/98</b> | Comparative study : 100% ; Collaborative study : 97.5%  |  |
| <b>Specify the standard test</b>  | Isolation + AGDIA Indirect-ELISA on pure culture (OEPP PM7/23)  |  |
| <b><u>Analytical specificity</u></b>  |   |  |
| <b>Specificity value</b>  | 100%  |  |
| <b>Number of strains/populations of target organisms tested</b>   | 50 (see attached downloadable file Appendix 1)  |  |
| <b>Number of non-target organisms tested</b>  | 53 (see attached downloadable file Appendix 2)  |  |
| <b>Cross reacts with (specify the species)</b>  | The restriction step performed after the N-PCR enables to exclude all the tested strains. Without the restriction step, a <i>Xanthomonas axonopodis</i> pv. <i>allii</i> strain and some <i>Xanthomonas campestris</i> pv. <i>syngonii</i> strains can not be excluded. |  |
| <b><u>Diagnostic Specificity</u></b>  |   |  |
| <b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>         | Comparative study : 96% ; Collaborative study : 95%   |  |
| <b>Specify the standard test</b>  | Isolation + AGDIA Indirect-ELISA on pure culture (OEPP PM7/23)  |  |
| <b><u>Reproducibility</u></b>   |   |  |
| <b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>  | 93%   |  |
| <b><u>Repeatability</u></b>   |   |  |
| <b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>  | 94%   |  |
| <b><u>Test performance study</u></b>  |   |  |
| <b>Test performance study?</b>  | Yes   |  |
| <b>Include brief details of the test</b>  | Results obtained with the N-PCR are excellent for all criteria  |  |

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| <p><b>performance study and its output. It available, provide a link to published article/report</b></p>     | <p>(&gt;= 90%) and not statistically different from results obtained with the standard test.<br/>The most important difference between the N-PCR and the standard test concerns the analytical sensitivity. Therefore, we proposed maintaining a step of pathogen isolation in the revised EPPO detection scheme.</p>   |
| <p><b><u>Other information</u></b></p>   |   |
| <p><b>Any other information considered useful<br/>e.g. robustness, ease of performing the test, etc.</b></p> | <p>When other criteria besides technical performance are considered, the N-PCR has advantages compared to the other methods tested :</p> <ul style="list-style-type: none"> <li>-It produces results more quickly (2-3 days) than the reference method for approximately the same cost.</li> <li>-It is easily transferable in comparison to isolation and the IF test, which require experience for recognising the typical bacteria.</li> </ul> |
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| <p>The following complementary files are available online:</p>   | <ul style="list-style-type: none"> <li>• <a href="#">Appendix_1-List_target_strains</a></li> <li>• <a href="#">Appendix_2-List_non_target_strains</a></li> <li>• <a href="#">Inter-laboratory ring test : Xanthomonas axonopodis pv. dieffenbachiae in Anthurium (Report Xad01-version 2)</a></li> </ul>  |