

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES**  
(11-17239)

**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Target Organism</b>	Xylella fastidiosa	
<b>Short description</b>	Detection of Xylella fastidiosa by real-time PCR in plant material (Harper et al., 2010, Erratum 2013)	
<b>Laboratory contact details</b>	Anses Plant Health Laboratory - Bacteriology, Virology and GMO Unit 7 rue Jean Dixméras, 49000 Angers, France	
<b>Date and reference of the validation report</b>	2015-09-30 - Rapport de caractérisation et de validation de méthode d'analyse - Détection de Xylella fastidiosa par PCR en temps réel sur plantes hôtes MA039	
<b>Validation process according to EPPO Standard PM 7/98:</b>	Yes	
<b>Reference of the test description</b>	0 - Harper et al., 2010, Erratum 2013 - MA 039 version 1 French reference method (www.anses.fr)	
<b>Is the test the same as described in the EPPO DP?</b>	Yes	
<b>Is the lab accredited for this test?</b>	No	
<b>Plant species tested (if relevant)</b>	Vitis vinifera, Citrus sinensis, Olea europaea	
<b>Matrices tested (if relevant)</b>	Petioles	
<b>List of methods used</b>		
<b>Method for extraction / isolation / baiting of target organism from matrix</b>	X	QuickPick™ Plant DNA kit (Bio-Nobile) Automated protocol with KingFisher™ mL (Thermo Scientific)
<b>Molecular methods, e.g. hybridization, PCR and real time PCR</b>	X	Real-time PCR Harper et al., 2010 (erratum 2013)
<b>Serological methods: IF, ELISA, Direct Tissue Blot Immuno Assay</b>		
<b>Plating methods: selective isolation</b>		
<b>Bioassay methods: selective enrichment in host plants, baiting, plant test and grafting.</b>		
<b>Pathogenicity test</b>		
<b>Fingerprint methods: protein</b>		

<b>profiling, fatty acid profiling &amp; DNA profiling</b>		
<b>Morphological and morphometrical methods intended for identification</b>		
<b>Biochemical methods: e.g. enzyme electrophoresis, protein profiling</b>		
<b>Other</b>		
<b>Analytical sensitivity (= limit of detection)</b>		
<b>What is smallest amount of target that can be detected reliably?</b>	<ul style="list-style-type: none"> <li>- Grapevine: <math>\approx 10^3</math> bact./mL</li> <li>- Orange tree: <math>\approx 10^2</math> bact./mL</li> <li>- Olive tree: <math>\approx 10^5</math> bact./mL</li> </ul> <p>With a probability of detection of 100%</p>	
<b>Diagnostic sensitivity</b>		
<b>Proportion of infected/infested samples tested positive compared to results from the standard test , see appendix 2 of PM 7/98</b>	<ul style="list-style-type: none"> <li>- Grapevine: 94%</li> <li>- Orange tree: 100%</li> <li>- Olive tree: 67%</li> </ul>	
<b>Specify the standard test</b>	<ul style="list-style-type: none"> <li>- Spiked matrices with bacterial concentration from <math>10^2</math> to <math>10^5</math> bact./mL</li> <li>- Grapevine spiked with <i>X. f. subsp. fastidiosa</i> (CFBP7970)</li> <li>- Orange tree spiked with <i>X. f. subsp. pauca</i> (CFBP8072)</li> <li>- Olive tree spiked with <i>X. f. subsp. multiplex</i> (CFBP8173)</li> </ul> <p>15 samples per matrix 30 DNA extraction per matrix 60 amplifications per matrix</p>	
<b>Analytical specificity</b>		
<b>Specificity value</b>	100%	
<b>Number of strains/populations of target organisms tested</b>	<p>Inclusivity tested with 19 target strains: 100%</p> <ul style="list-style-type: none"> <li>- <i>X.f. subsp. fastidiosa</i> (CFBP8069 - LSV 0056 / CFBP7970 - LSV 2434 / CFBP8082 - LSV 4040 / CFBP8071 - LSV 4041 / CFBP8083 - LSV 4042 / CFBP8073-LSV4209 / CFBP8351 - LSV4626)</li> <li>- <i>X.f. subsp. pauca</i> (CFBP8072 - LSV 4103)</li> <li>- <i>X.f. subsp. sandyi</i> (CFBP8077 - LSV 4236 / CFBP 8356 - LSV4627 / LSV4628 / LSV4639 / LSV4659)</li> <li>- <i>X.f. subsp. multiplex</i> (CFBP8068 - LSV 0054 / CFBP8070 - LSV 4038/ CFBP8173 - LSV 4039 / CFBP8075 - LSV 4230/ CFBP8076 - LSV 4231 / CFBP8078 - LSV 4311)</li> </ul> <p>Bacterial suspension concentration of about <math>10^7</math> bact./mL</p>	
<b>Number of non-target organisms tested</b>	<p>Exclusivity tested with 29 non-target strains: 100%</p> <ul style="list-style-type: none"> <li>- 1 <i>Xylophilus ampelinus</i> (CFBP2098)</li> <li>- 2 <i>Xanthomonas arboricola</i> pv. <i>pruni</i> (LSV2574/LSV 2573)</li> <li>- 1 <i>Xanthomonas arboricola</i> pv. <i>juglandis</i> (LSV0862)</li> <li>- 1 <i>Xanthomonas axonopodis</i> pv. <i>citri</i> (LSV2647)</li> <li>- 1 <i>Xanthomonas axonopodis</i> pv. <i>aurantifolia</i> (LSV2680)</li> <li>- 2 <i>Xanthomonas axonopodis</i> pv. <i>phaseoli</i> (LSV1014/LSV3161)</li> <li>- 1 <i>Xanthomonas axonopodis</i> pv. <i>fragariae</i> (LSV3151)</li> </ul>	

	<ul style="list-style-type: none"> <li>- 1 Xanthomonas fragariae (LSV2553)</li> <li>- 1 Xanthomonas hortorum pv. carotae (LSV1776)</li> <li>- 1 Xanthomonas campestris pv. campestris (LSV0455)</li> <li>- 1 Xanthomonas campestris pv. juglandis (LSV1158)</li> <li>- 1 Xanthomonas hortorum pv. hederata (LSV2303)</li> <li>- 1 Xanthomonas translucens pv. graminis (LSV0628)</li> <li>- 1 Xanthomonas translucens pv. hordei (LSV0629)</li> <li>- 1 Xanthomonas oryzae pv. oryzae (LSV0865)</li> <li>- 1 Ca. Liberibacter asiaticus</li> <li>- 1 Ca. L. africanus</li> <li>- 6 saprophytic bacteria saprophytes isolated from Coffea spp.</li> <li>- 4 bactéries saprophytes isolées de Citrus sinensis</li> </ul> <p>Bacterial suspension concentration of about <math>10^7</math> bact./mL</p>
<b>Cross reacts with (specify the species)</b>	None
<b>Diagnostic Specificity</b>	
<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>	<ul style="list-style-type: none"> <li>- Grapevine: 100%</li> <li>- Orange tree: 100%</li> <li>- Olive tree: 100%</li> </ul>
<b>Specify the standard test</b>	<p>Spiked matrices with bacterial concentration from <math>10^3</math> to <math>10^5</math> bact./mL</p> <ul style="list-style-type: none"> <li>- Grapevine spiked with X. f. subsp. fastidiosa (CFBP7970)</li> <li>- Orange tree spiked with X. f. subsp. pauca (CFBP8072)</li> <li>- Olive tree spiked with X. f. subsp. multiplex (CFBP8173)</li> </ul> <p>15 samples per matrix 30 DNA extraction per matrix 60 amplifications per matrix</p>
<b>Reproducibility</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	98%
<b>Repeatability</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	<ul style="list-style-type: none"> <li>- Grapevine: 96%</li> <li>- Orange tree: 100%</li> <li>- Olive tree: 100%</li> </ul>
<b>Test performance study</b>	
<b>Test performance study?</b>	Yes
<b>Include brief details of the test performance study and its output. If available, provide a link to published article/report</b>	<p>A test performance study was performed in 2014 for the Real time PCR Harper et al., 2010 method but with another DNA extraction method (DNeasy® Plant mini kit (Qiagen))</p> <p>Analytical sensitivity (with a probability of detection of 100%):</p> <ul style="list-style-type: none"> <li>- Orange tree: <math>\approx 10^2</math> bact./mL</li> <li>- Grapevine: <math>\approx 10^6</math> bact./mL</li> <li>- Peach tree: <math>\approx 10^4</math> bact./mL</li> <li>- Olive tree: <math>\approx 10^5</math> bact./mL</li> <li>- Coffee tree: <math>\approx 10^4</math> bact./mL</li> </ul> <p>Diagnostic sensitivity: 97%</p>

	Diagnostic specificity: 97% Reproducibility: 84% Repeatability: 91%
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<b><u>Other information</u></b>	
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<b>Any other information considered useful e.g. robustness, ease of performing the test, etc.</b>	
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For information, a proficiency test was performed in 2015 for this method.
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