

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES**  
**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Laboratory contact details</b>	Netherlands Institute for Vectors, Invasive plants and Plant health P.O. Box 9102, 6700 HC Wageningen, Netherlands
<b>Short description of the test</b>	Detection and identification of <i>Pantoea stewartii</i> subsp. <i>stewartii</i> with real-time PCR
<b>Date, reference of the validation report</b>	2022-04-28 - 2022.molbio.003
<b>Validation process according to EPPO Standard PM7/98?</b>	yes
<b>Is the lab accredited for this test?</b>	yes
<b>Was the validated data generated in the framework of a project?</b>	Other_project
<b>If yes, please specify</b>	Dutch phytosanitary project
<b>Description of the test</b>	
<b>Organism(s)</b>	<i>Pantoea stewartii</i> subsp. <i>stewartii</i> (ERWIST)
<b>Detection / identification</b>	detection and identification
<b>Method(s)</b>	Molecular real time RT PCR
<b>Method: Molecular real time RT PCR</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an EPPO diagnostic protocol</b>	yes
<b>New test being considered for inclusion in the next version of the EPPO diagnostic protocol?</b>	no
<b>EPPO Diagnostic Protocol name</b>	PM 7/060 <i>Pantoea stewartii</i> (version 2)
<b>As or adapted from an IPPC diagnostic protocol</b>	no
<b>Is the test modified compared to the reference test</b>	no
<b>Kit</b>	
<b>Is a kit used</b>	no
<b>Other information</b>	
<b>Reaction type</b>	Simplex
<b>Other details on the test</b>	The Pal et al. real-time PCR was run in 20 µL

	including 4 µL of sample using 10 µL of GoTaq qPCR mastermix by Promega with the following PCR protocol: 10 minutes at 95°C followed by 40 cycles of 95°C for 15 seconds and 66°C for 1 minute. Subsequently, a melt curve was produced: An increase from 65°C to 95°C in increments of 0.5°C per 10 seconds. Primers were used in accordance with the original protocol (Table 1). The reaction mix was finished off with DNase/RNase free water. Primers were obtained from Eurogentec. The real-time protocol was run on the CFX96 Touch Real-Time PCR Detection System. Threshold values were automatically calculated for each run. Table 1: Primers used during this validation Oligo name Sequence (5' - 3') Final concentration cpsAB2313F AGA AAA CGC TGA TGC CAG AC 400 nM cpsR ACT ATC CTG ACT CAG GCA CT 400 n
<b>Performance Criteria :</b>	
<b>Organism 1.:</b>	<b><i>Pantoea stewartii</i> subsp. <i>stewartii</i>(ERWIST)</b>
<b><u>Analytical sensitivity</u></b>	
<b>What is smallest amount of target that can be detected reliably?</b>	12.14 CFU / reaction or 3035 CFU / mL
<b><u>Diagnostic sensitivity</u></b>	
<b>Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98</b>	100% The host matrix did not influence the performance of the test.
<b>Standard test(s)</b>	Two different sweet corn varieties were used (Tasty Sweet F1 and Jubilee). The status of the samples was known.
<b><u>Analytical specificity - inclusivity</u></b>	
<b>Number of strains/populations of target organisms tested</b>	13 isolates of Pss
<b>Specificity value</b>	100%
<b><u>Analytical specificity - exclusivity</u></b>	
<b>Number of non-target organisms tested</b>	Clavibacter nebraskensis (1x) Dickeya zea (2x) P. agglomerans pv. milletae (1x) P. alli (1x) P. ananatis (6x) P. ananatis pv. anantis (1x) P. ananatis pv. uredovora (1x) P. stewartii subsp. indologenes (6x) Paraburkholderia andropogonis (1x) Pseudomonas syringae pv. syringae (2x) Pseudomonas syringae pv. lapsa (1xc) Xanthomonas campestris pv. vasculorum (1x)
<b>Specificity value</b>	100%
<b><u>Diagnostic Specificity</u></b>	
<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>	100%
<b>Specify the test(s)</b>	Phosphate buffer was used as a negative control.

<b>Reproducibility</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% Three samples were isolated and tested by all three technicians.
<b>Repeatability</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% Three samples were isolated and tested in duplicate by all three technicians.
<b>Test performance study</b>	
<b>Test performance study?</b>	no
The following complementary files are available online:	
	<ul style="list-style-type: none"> <li>• <a href="#">Real-time PCR Pss</a></li> </ul>

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