

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION  
 ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES  
 Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Laboratory contact details</b>	Netherlands Institute for Vectors, Invasive plants and Plant health P.O. Box 9102, 6700 HC Wageningen, Netherlands
<b>Short description of the test</b>	Development and validation of a real-time RT-PCR assay for generic detection of Pospiviroids
<b>Date, reference of the validation report</b>	2012-04-04 - NRC-ref: 2010.molbio.015
<b>Validation process according to EPPO Standard PM7/98?</b>	yes
<b>Is the lab accredited for this test?</b>	no
<b>Was the validated data generated in the framework of a project?</b>	
<b>Description of the test</b>	
<b>Organism(s)</b>	Pospiviroid (1POSPG)
<b>Detection / identification</b>	detection
<b>Method(s)</b>	Molecular Extraction DNA RNA Molecular real time RT PCR
<b>Method: Molecular Extraction DNA RNA</b>	
<b>Reference of the test description</b>	
<b>Kit</b>	
<b>Is a kit used</b>	yes
<b>Manufacturer name</b>	QIAGEN
<b>Specify the kit used</b>	RNeasy Plant Mini Kit
<b>Kit used following the manufacturer's instructions?</b>	
<b>Other information</b>	
<b>Method: Molecular real time RT PCR</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an EPPO diagnostic protocol</b>	no
<b>As or adapted from an IPPC diagnostic protocol</b>	no
<b>Reference of the test</b>	M. Botermans, B.T.L.H. van de Vossenbergh, J.Th.J. Verhoeven, J.W. Roenhorst, M. Hooftman, R.

	Dekter, E.T.M. Meekes (2013) Development and validation of a real-time RT-PCR assay for generic detection of Pospiviroids. Journal of Virological Methods 187 43- 50
<b>Other information</b>	
<b>Are the performance characteristics included in the EPPO diagnostic protocol?</b>	<b>no</b>
<b>Performance Criteria :</b>	
<b>Organism 1.:</b>	<b>Pospiviroid(1POSPG)</b>
<b>Analytical sensitivity</b>	
<b>What is smallest amount of target that can be detected reliably?</b>	Undiluted infected leaf sap was considered 100% infected. Starting with undiluted plant sap from infected hosts up to 10 <sup>7</sup> times dilutes in sap of healthy tomato leaves, all pospiviroids species were detected up to a relative infection rate of 0.13% (=770 times diluted)
<b>Diagnostic sensitivity</b>	
<b>Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98</b>	1
<b>Standard test(s)</b>	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
<b>Analytical specificity - inclusivity</b>	
<b>Number of strains/populations of target organisms tested</b>	28 pospiviroid strains, see table 1 Botermans et al. (2013)
<b>Specificity value</b>	1
<b>Analytical specificity - exclusivity</b>	
<b>Number of non-target organisms tested</b>	8 non -ospiviroid strains and 8 tomato viruses, see table 1 Botermans et al. (2013)
<b>Specificity value</b>	
<b>Diagnostic Specificity</b>	
<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>	1
<b>Specify the test(s)</b>	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
<b>Reproducibility</b>	

<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% at undiluted, 500 x, and 1000x diluted
<b>Repeatability</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% at undiluted, 500 x, and 1000x diluted
<b>Test performance study</b>	
<b>Test performance study?</b>	yes
<b>Brief details of the test performance study and its output. It available, link to published article/report</b>	Three laboratories participated in the inter-laboratory comparison: Dutch General Inspection Service for Agricultural Seed and Seed Potatoes (NAK, Emmeloord), Naktuinbouw (Roelofarendsveen) and the National Reference Centre of the National Plant Protection Organization (Wageningen). Sixteen samples of tomato leaves infected with PSTVd, TASVd or Tomato chlorotic dwarf viroid (TCDVd) at different relative infection rates were tested at the three laboratories (Table 6, Botermans et al. 2013)
<b>Other information</b>	
<b>Any other information considered useful</b>	For tomato leaves the assay is not influenced by the matrix (selectivity). To determine the robustness of the test an alternative RNA extraction kit was used: the Sbeadex maxi plant kit on a Kingfisher KF96 system, and Real-time RT-PCRs were carried out on different real-time PCR machines. The test results were similar for all samples tested.
The following complementary files are available online:	<ul style="list-style-type: none"> <li>• <a href="#">Development and validation of a real-time RT-PCR assay for generic detection of pospiviroids</a></li> </ul>

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