

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION  
ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES  
Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Laboratory contact details</b>	Netherlands Institute for Vectors, Invasive plants and Plant health P.O. Box 9102, 6700 HC Wageningen, Netherlands
<b>Short description of the test</b>	Validation of a conventional RT-PCR test for detection and identification of PSTVd, TCDVd, MPVd and TPMVd using primers 2H1/3H1 described by Shamoul et al. (1997)
<b>Date, reference of the validation report</b>	2013-09-17 - NRC-ref: 2011.molbio.006
<b>Validation process according to EPPO Standard PM7/98?</b>	yes
<b>Is the lab accredited for this test?</b>	no
<b>Was the validated data generated in the framework of a project?</b>	
<b>Description of the test</b>	
<b>Organism(s)</b>	Pospiviroid chloronani(TCDVD0) Pospiviroid fusituberis(PSTVD0)
<b>Detection / identification</b>	detection and identification
<b>Method(s)</b>	Molecular Extraction DNA RNA Molecular Conventional RT PCR Molecular other
<b>Method: Molecular Extraction DNA RNA</b>	
<b>Reference of the test description</b>	
<b>Kit</b>	
<b>Is a kit used</b>	yes
<b>Manufacturer name</b>	QIAGEN
<b>Specify the kit used</b>	RNeasy Plant Mini Kit
Kit used following the manufacturer's instructions?	
<b>Other information</b>	
<b>Method: Molecular Conventional RT PCR</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an IPPC diagnostic protocol</b>	yes
<b>IPPC diagnostic Protocol name</b>	ISPM 27 Annex 07 DP 07: Potato spindle tuber

	viroid (version 2006)
<b>Name of the test</b>	Conventional RT-PCR using the primers of Shamloul et al. (1997)
<b>Other information</b>	
<b>Other details on the test</b>	section 3.3.4.1 Conventional RT-PCR using the primers of Shamloul et al. (1997) Shamloul AM, Hadidi A, Zhu SF, Singh RP, Sagredo B (1997) Sensitive detection of potato spindle tuber viroid using RT-PCR and identification of a viroid variant naturally infecting pepino plants. Can. J. Pl. Pathol. 19: 89-96 RT-PCR by 2H1/3H1 primers and sequencing of the amplicon
<b>Method: Molecular other</b>	
<b>Reference of the test description</b>	
<b>Other information</b>	
<b>Other details on the test</b>	Sequencing
<b>Are the performance characteristics included in the EPPO diagnostic protocol?</b>	<b>no</b>
<b>Performance Criteria :</b>	
<b>Organism 1.:</b>	<b>Pospiviroid chloronani(TCDVD0)</b>
<b>Analytical sensitivity</b>	
<b>What is smallest amount of target that can be detected reliably?</b>	Undiluted leaf sap was considered 100% infected. Starting with undiluted plant sap from PSTVd and TCDVd infected hosts up to 10 <sup>7</sup> times dilutes in sap of healthy tomato leaves, all PSTVd and TCDVd strains were detected up to a relative infection rate of 10% (=10 times diluted) .
<b>Diagnostic sensitivity</b>	
<b>Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98</b>	1
<b>Standard test(s)</b>	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
<b>Analytical specificity - inclusivity</b>	
<b>Number of strains/populations of target organisms tested</b>	7 pospiviroid strains, see table 4 validation report
<b>Specificity value</b>	1
<b>Analytical specificity - exclusivity</b>	
<b>Number of non-target organisms tested</b>	10 (4 Avsunviroidae and 6 Pospiviroidae strains), see table 6 validation report
<b>Specificity value</b>	

<b>Diagnostic Specificity</b>	
<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>	1
<b>Specify the test(s)</b>	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
<b>Reproducibility</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% at undiluted, 2x and 4x diluted
<b>Repeatability</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% at undiluted, 2x and 4x diluted
<b>Organism 2.:</b>	<b>Pospiviroid fusituberis(PSTVD0)</b>
<b>Analytical sensitivity</b>	
<b>What is smallest amount of target that can be detected reliably?</b>	Undiluted leaf sap was considered 100% infected. Starting with undiluted plant sap from PSTVd and TCDVd infected hosts up to 10 <sup>7</sup> times dilutes in sap of healthy tomato leaves, all PSTVd and TCDVd strains were detected up to a relative infection rate of 10% (=10 times diluted) .
<b>Diagnostic sensitivity</b>	
<b>Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98</b>	1
<b>Standard test(s)</b>	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
<b>Analytical specificity - inclusivity</b>	
<b>Number of strains/populations of target organisms tested</b>	7 pospiviroid strains, see table 4 validation report
<b>Specificity value</b>	1
<b>Analytical specificity - exclusivity</b>	
<b>Number of non-target organisms tested</b>	10 (4 Avsunviroidae and 6 Pospiviroidae strains), see table 6 validation report
<b>Specificity value</b>	
<b>Diagnostic Specificity</b>	
<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared</b>	1

<b>to results from a standard test</b>	
<b>Specify the test(s)</b>	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
<b>Reproducibility</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% at undiluted, 2x and 4x diluted
<b>Repeatability</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% at undiluted, 2x and 4x diluted
<b>Test performance study</b>	
<b>Test performance study?</b>	no
<b>Other information</b>	
<b>Any other information considered useful</b>	To determine the robustness of the test two different reaction mixes were used to generate PCR products: OneStep RT-PCR kit (Qiagen) and SuperScript One-Step RT-PCR System with Platinum Taq (Invitrogen). The latter yielded the best results. Different extraction buffers were used. Using a GH plus buffer with a heating step yielded the best results. Freezing the samples prior to analysis did not influence the qualitative results obtained.
The following complementary files are available online:	<ul style="list-style-type: none"> <li>• <a href="#">Validation of a conventional RT-PCR test for detection and identification of PSTVd, TCDVd, MPVd and TPMVd using primers described by Shamoul et al. (1997). (In Dutch)</a></li> </ul>

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