

EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION
ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES
Summary sheet of validation data for a diagnostic test

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

Laboratory contact details	National Reference Centre, National Plant Protection Organization P.O. Box 9102, 6700 HC Wageningen, Netherlands
Short description of the test	Validation of a conventional RT-PCR test for detection and identification of PSTVd, TCDVd, MPVd and TPMVd using primers 2H1/3H1 described by Shamoul et al. (1997)
Date, reference of the validation report	2013-09-17 - NRC-ref: 2011.molbio.006
Validation process according to EPPO Standard PM7/98?	yes
Is the lab accredited for this test?	no
Was the validated data generated in the framework of a project?	
If yes, please specify	
Description of the test	
Organism(s)	Tomato chlorotic dwarf viroid(TCDVD0) Potato spindle tuber viroid(PSTVD0)
Detection / identification	detection and identification
Matrix(ces) tested	Leaves leaves
Plant species tested	Solanum lycopersicum
Method(s)	Molecular Extraction DNA RNA Molecular Conventional RT PCR Molecular other
Method: Molecular Extraction DNA RNA	
Reference of the test description	
As or adapted from an EPPO diagnostic protocol	
New test being considered for inclusion in the next version of the EPPO diagnostic protocol?	
As or adapted from an IPPC diagnostic protocol	
Is the test modified compared to the reference test	
Kit	

Is a kit used	yes
Manufacturer name	QIAGEN
Specify the kit used	RNeasy Plant Mini Kit
Kit used following the manufacturer's instructions?	
Other information	
Other details on the test	
Method: Molecular Conventional RT PCR	
Reference of the test description	
As or adapted from an EPPO diagnostic protocol	
New test being considered for inclusion in the next version of the EPPO diagnostic protocol?	
As or adapted from an IPPC diagnostic protocol	yes
IPPC diagnostic Protocol name	ISPM 27 Annex 07 DP 07: Potato spindle tuber viroid (version 2006)
Name of the test	Conventional RT-PCR using the primers of Shamloul et al. (1997)
Is the test modified compared to the reference test	
Kit	
Is a kit used	
Other information	
Reaction type	
Other details on the test	section 3.3.4.1 Conventional RT-PCR using the primers of Shamloul et al. (1997) Shamloul AM, Hadidi A, Zhu SF, Singh RP, Sagredo B (1997) Sensitive detection of potato spindle tuber viroid using RT-PCR and identification of a viroid variant naturally infecting pepino plants. Can. J. Pl. Pathol. 19: 89-96 RT-PCR by 2H1/3H1 primers and sequencing of the amplicon
Method: Molecular other	
Reference of the test description	
As or adapted from an EPPO diagnostic protocol	
New test being considered for inclusion in the next version of the EPPO diagnostic protocol?	
As or adapted from an IPPC diagnostic protocol	
Is the test modified compared to the reference test	
Kit	

Is a kit used	
Other information	
Reaction type	
Other details on the test	Sequencing
Are the performance characteristics included in the EPPO diagnostic protocol?	no
Performance Criteria :	
Organism 1.:	Tomato chlorotic dwarf viroid(TCDVD0)
Analytical sensitivity	
What is smallest amount of target that can be detected reliably?	Undiluted leaf sap was considered 100% infected. Starting with undiluted plant sap from PSTVd and TCDVd infected hosts up to 10 ⁷ times dilutes in sap of healthy tomato leaves, all PSTVd and TCDVd strains were detected up to a relative infection rate of 10% (=10 times diluted) .
Diagnostic sensitivity	
Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98	1
Standard test(s)	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
Analytical specificity - inclusivity	
Number of strains/populations of target organisms tested	7 pospiviroid strains, see table 4 validation report
Specificity value	1
Analytical specificity - exclusivity	
Number of non-target organisms tested	10 (4 Avsunviroidae and 6 Pospiviroidae strains), see table 6 validation report
Specificity value	
Cross reacts with	
Diagnostic Specificity	
Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test	1
Specify the test(s)	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.

Reproducibility	
Provide the calculated % of agreement for a given level of the pest (see PM 7/98)	100% at undiluted, 2x and 4x diluted
Repeatability	
Provide the calculated % of agreement for a given level of the pest (see PM 7/98)	100% at undiluted, 2x and 4x diluted
Organism 2.:	Potato spindle tuber viroid(PSTVD0)
Analytical sensitivity	
What is smallest amount of target that can be detected reliably?	Undiluted leaf sap was considered 100% infected. Starting with undiluted plant sap from PSTVd and TCDVd infected hosts up to 10 ⁷ times dilutes in sap of healthy tomato leaves, all PSTVd and TCDVd strains were detected up to a relative infection rate of 10% (=10 times diluted) .
Diagnostic sensitivity	
Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98	1
Standard test(s)	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
Analytical specificity - inclusivity	
Number of strains/populations of target organisms tested	7 pospiviroid strains, see table 4 validation report
Specificity value	1
Analytical specificity - exclusivity	
Number of non-target organisms tested	10 (4 Avsunviroidae and 6 Pospiviroidae strains), see table 6 validation report
Specificity value	
Cross reacts with	
Diagnostic Specificity	
Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test	1
Specify the test(s)	PCR sequencing of the complete viroid genome is considered the standard test. This can be achieved with primers Vid-FW/Vid-RE (CLVd, PSTVd and TCDVd) and 3H1/2H1 (PSTVd, TCDVd, MPVd and TPMVd). For other pospiviroids primers Posp1-FW/Posp1-RE are used to sequence the partial viroid genome.
Reproducibility	

Provide the calculated % of agreement for a given level of the pest (see PM 7/98)	100% at undilluted, 2x and 4x diluted
Repeatability	
Provide the calculated % of agreement for a given level of the pest (see PM 7/98)	100% at undilluted, 2x and 4x diluted
Test performance study	
Test performance study?	no
Brief details of the test performance study and its output. It available, link to published article/report	
Other information	
Any other information considered useful	To determine the robustness of the test two different reaction mixes were used to generate PCR products: OneStep RT-PCR kit (Qiagen) and SuperScript One-Step RT-PCR System with Platinum Taq (Invitrogen). The latter yielded the best results. Different extraction buffers were used. Using a GH plus buffer with a heating step yielded the best results. Freezing the samples prior to analysis did not influence the qualitative results obtained.
The following complementary files are available online:	<ul style="list-style-type: none"> • Validation of a conventional RT-PCR test for detection and identification of PSTVd, TCDVd, MPVd and TPMVd using primers described by Shamoul et al. (1997). (In Dutch)

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