

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES**  
**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Laboratory contact details</b>	Bacteriology. Instituto Valenciano de Investigaciones Agrarias CV-315, km. 10.7, 46113 Moncada, Spain
<b>Short description of the test</b>	Analytical sensitivity of the detection of 'Candidatus Liberibacter solanacearum' by real time PCR in carrot seeds using Plant Print diagnostic kit
<b>Date, reference of the validation report</b>	2016-05-06 - Validation assay December 2015 - PNT 14 Validacion Xcc y CaLSol_REV 3 / 2015-2
<b>Validation process according to EPPO Standard PM7/98?</b>	yes
<b>Is the lab accredited for this test?</b>	yes
<b>Was the validated data generated in the framework of a project?</b>	
<b>Description of the test</b>	
<b>Organism(s)</b>	'Candidatus Liberibacter solanacearum' (LIBEPS)
<b>Detection / identification</b>	detection
<b>Method(s)</b>	Extraction Molecular real time PCR
<b>Method: Extraction</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an EPPO diagnostic protocol</b>	no
<b>As or adapted from an IPPC diagnostic protocol</b>	no
<b>Reference of the test</b>	Teresani, GR, Bertolini, E., Alfaro-Fernandez, A. et al. 2014 Association of 'Candidatus Liberibacter solanacearum' with a Vegetative Disorder of Celery in Spain and Development of a Real-Time PCR Method for Its Detection. <a href="http://dx.doi.org/10.1094/PHYTO-07-13-0182-R">http://dx.doi.org/10.1094/PHYTO-07-13-0182-R</a> E. Bertolini et al. Transmission of 'Candidatus Liberibacter solanacearum' in carrot seeds. Plant Pathology 2014. Doi:10.1111/ppa.12245
<b>Other information</b>	
<b>Other details on the test</b>	Direct sample preparation without DNA purification

	(spot procedure) according to Bertolini et al. 2014, Teresani et al. 2014
<b>Method: Molecular real time PCR</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an EPPO diagnostic protocol</b>	yes
<b>EPPO Diagnostic Protocol name</b>	PM 7/143 ' <i>Candidatus Liberibacter solanacearum</i> ' (version 1)
<b>Name of the test</b>	Real-time PCR based on 16S rRNA gene (Teresani et al., 2014)
<b>Is the test modified compared to the reference test</b>	yes Use of a kit
<b>Kit</b>	
<b>Is a kit used</b>	yes
<b>Manufacturer name</b>	PLANT PRINT
<b>Specify the kit used</b>	'Candidatus Liberibacter solanacearum' Complete real-time PCR kit for direct screening (Ref: CaLsol/100)
Kit used following the manufacturer's instructions?	
<b>Other information</b>	
<b>Other details on the test</b>	Real time PCR using Plant Print diagnostics kit, based on Bertolini et al. 2014, Teresani et al 2014
<b>Are the performance characteristics included in the EPPO diagnostic protocol?</b>	<b>no</b>
<b>Performance Criteria :</b>	
<b>Organism 1.:</b>	<b>'Candidatus Liberibacter solanacearum'(LIBEPS)</b>
<b>Analytical sensitivity</b>	
<b>What is smallest amount of target that can be detected reliably?</b>	Detection of 100% (12/12) dilution 1/1000 (-3) of a spiked sample extract of carrot seeds spiked with an approximate concentration of 1,7 nanograms/microlitre of purified DNA. (Approx. 10-100 cells by direct sample preparation according calculation of Bertolini et al. 2014.) Detection of 75% (9/12) dilution 1/10000 (-4) of the same spiked samples by the standard test (standard test was the same real-time PCR performed with a previous extraction of DNA by CTAB protocol) (Approx. 3-100 cells by direct sample preparation according calculation of Bertolini et al. 2014.)
<b>Diagnostic sensitivity</b>	
<b>Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98</b>	60% for the spot test 76% for the standard test (standard test was the same real-time PCR performed with a previous extraction of DNA by CTAB protocol)

<b>Standard test(s)</b>	-60 spiked samples of -1, -2, -3, -4 and -5 dilutions of a spiked sample extract of seeds spiked with purified DNA from naturally contaminated seeds -36 samples positive /60 using the spot method -46 samples positive /60 using the standard test (performed with a previous extraction of DNA by CTAB protocol)
<b>Reproducibility</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% The reproducibility was calculated performing three series of dilutions of spiked samples (-1 to -5 dilution) analyzed in different days.
<b>Repeatability</b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	100% The repeatability was calculated performing the analysis of three series of spiked samples (-1 to -5 dilution) and three replicates/serie.
<b>Test performance study</b>	
<b>Test performance study?</b>	no
<b>Other information</b>	
<b>Any other information considered useful</b>	The method of analysis of carrot seeds with the diagnostic kit evaluated is simple to use, rapid and accurate. It can be applied for rapid testing of large number of samples of carrot seeds. For maximum accuracy a previous CTAB extraction or other types of DNA extraction is advised.
The following complementary files are available online:	
	<ul style="list-style-type: none"> <li>• <a href="#">Ejercicio control interno CaLsol y Xcc 2015_2</a></li> </ul>

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