EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION
ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES
Summary sheet of validation data for a diagnostic test

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

Laboratory contact details	Council for Agricultural Research and Economics–Research Centre for Plant Protection and Certification Via Carlo Giuseppe Bertero, 22, 00156 Rome, Italy
Short description of the test	Detection of 'Candidatus Phytoplasma mali' by direct PCR
Date, reference of the validation report	2013-01-01 - 97 ; 1) www.strateco.it 2)Pasquini et al., 2013. Petria 23(3),461-490
Validation process according to EPPO Standard PM7/98?	yes
Is the lab accredited for this test?	no
Was the validated data generated in the framework of a project?	Other_project
If yes, please specify	Project financed by the Italian Ministry of Agriculture (ARNADIA)
Description of the test	
Organism(s)	'Candidatus Phytoplasma mali' (PHYPMA)
Detection / identification	detection
Method(s)	Molecular Extraction DNA RNA Molecular Conventional PCR
Method: Molecular Extraction DNA RNA	
Reference of the test description	
Kit	
Is a kit used	yes
Manufacturer name	QIAGEN
Specify the kit used	DNeasy Plant Mini Kit
Kit used following the manufacturer's instructions?	
Other information	
Other details on the test	Commercial kit (DNeasy Plant Mini kit Qiagen) from leaf midribs or phloem tissue, previously powdered with liquid nitrogen. An alternative protocol has been used in the case of not availability of liquid nitrogen for the initial powdering of plant material. (Pasquini et al., 2013) Pasquini G., Ferretti L.,

	Bertaccini A., Bianco P.A., Casati P., Costantini E., Martini M., Marzachì C., Palmano S., Paltrinieri S., 2013. Protocollo diagnostico per 'Candidatus Phytoplasma mali' (AP). Petria 23 (3), 461-490
Method: Molecular Conventional PCR	
Reference of the test description	
As or adapted from an EPPO diagnostic protocol	no
As or adapted from an IPPC diagnostic protocol	no
Reference of the test	Smart CD., B. Schneider, CL. Blomquist, LJ. Guerra, NA. Harrison, U. Ahrens, KH. Lorenz, E. Seemuller, BC. Kirkpatrick, 1996. Phytoplasma-specific PCR primers based on the sequences of the 16S-23S rRNA spacer region. Applied and Environmental Microbiology, 42, 2988-2993
Other information	
Other details on the test	PCR with primers fAT/rAS (Smart et al.,1996) specific for 16Sr-XA and XC phytoplasmas
Are the performance characteristics included in the EPPO diagnostic protocol?	**no**
Performance Criteria :	
Organism 1.:	**'Candidatus Phytoplasma mali'(PHYPMA)**
Analytical sensitivity	
What is smallest amount of target that can be detected reliably?	The analytical sensitivity was calculated analyzing three samples at seven diluition levels (1/1-1/1.000.000). The dilutions were in DNA from an healthy apple sample. Last dilution level with 100% positive results for all three samples: 1/100
Diagnostic sensitivity	
Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98	21 'target' samples: 20 apple samples infected by 'Ca. P. mali' coming from different Italian areas and one sample of pear infected by 'Candidatus Phytoplasma pyri' (PD phytoplasma) Diagnostic sensitivity: 81% Six laboratories were involved in performing this method within the ringtest. The diagnostic sensitivity was determined by using 20 apple samples positive for 'Ca. P. mali', coming from different Italian areas. Within the ringtest 3 diagnostic methods were compared. Diagnostic sensitivity: 81%
Standard test(s)	Other methodologies included in the ringtest: - Direct universal PCR with primers P1/16S-Sr, followed by nested 16SrX group specific primers fO1/rO1 (Lorenz et al., 1995) - SYBR Green real time PCR (Galetto et al., 2005)
Analytical specificity - inclusivity	
Number of strains/populations of target organisms tested	21 'target' samples: 20 apple samples infected by 'Ca. P. mali' coming from different Italian areas and

	one sample of pear infected by 'Candidatus Phytoplasma pyri' (PD phytoplasma)
Specificity value	Analytical specificity: 100%
Analytical specificity - exclusivity	
Number of non-target organisms tested	Four 'non target' samples were included: phytobacteria commonly spread on pome fruits and ESFY phytoplasma belonging to 16SrX group: - Extracted DNA from a pear infected by Pseudomonas syringae pv. syringae - Extracted DNA from an apple infected by Erwinia amilovora - two samples of plums infected by 'Candidatus Phytoplasma prunorum' (ESFY phytoplasma)
Specificity value	Analytical specificity: 100%. No cross reaction
Diagnostic Specificity	
Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test	Five samples of apple tree uninfected (certified material) Diagnostic specificity: 100%
Specify the test(s)	Other methodologies included in the ringtest: - Direct universal PCR with primers P1/16S-Sr, followed by nested 16SrX group specific primers fO1/rO1 (Lorenz et al., 1995) - SYBR Green real time PCR (Galetto et al., 2005)
Reproducibility	
Provide the calculated % of agreement for a given level of the pest (see PM 7/98)	The reproducibility was calculated analyzing in six laboratories all samples included in diagnostic specificity and sensitivity tests. Reproducibility: 89.1%
Repeatability	
Provide the calculated % of agreement for a given level of the pest (see PM 7/98)	The repeatability was calculated in three laboratories analyzing three samples at seven diluition levels (1/1-1/1.000.000). The dilutions were in DNA from an healthy apple sample. Repeatability: 100%
Test performance study	
Test performance study?	yes
Brief details of the test performance study and its output.It available, link to published article/report	A ringtest was organized with the official Italian phytosanitary laboratories within a Project financed by the Italian Ministry of Agriculture (ARNADIA) for the definition of 'Italian reference protocols'.

Creation date: 2015-02-11 00:00:00 - Last update: 2020-10-08 18:14:15