

**EUROPEAN AND MEDITERRANEAN PLANT PROTECTION ORGANIZATION**  
**ORGANISATION EUROPEENNE ET MEDITERRANEENNE POUR LA PROTECTION DES PLANTES**  
**Summary sheet of validation data for a diagnostic test**

The EPPO Standard PM 7/98 *Specific requirements for laboratories preparing accreditation for a plant pest diagnostic activity* describes how validation should be conducted. It also includes definitions of performance criteria.

<b>Laboratory contact details</b>	Council for Agricultural Research and Economics- Research Centre for Plant Protection and Certification Via Carlo Giuseppe Bertero, 22, 00156 Rome, Italy
<b>Short description of the test</b>	Detection of 'Candidatus Phytoplasma mali' by direct and nested PCR
<b>Date, reference of the validation report</b>	2013-01-01 - 1) www. strateco.it 2) Pasquini et al., Petria 23(3), 461-490
<b>Validation process according to EPPO Standard PM7/98?</b>	yes
<b>Is the lab accredited for this test?</b>	no
<b>Was the validated data generated in the framework of a project?</b>	Other_project
<b>If yes, please specify</b>	Project financed by the Italian Ministry of Agriculture (ARNADIA) for the definition of 'Italian reference protocols'.
<b>Description of the test</b>	
<b>Organism(s)</b>	'Candidatus Phytoplasma mali' (PHYPPMA)
<b>Detection / identification</b>	detection
<b>Method(s)</b>	Molecular Extraction DNA RNA Molecular Conventional PCR
<b>Method: Molecular Extraction DNA RNA</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an EPPO diagnostic protocol</b>	no
<b>Kit</b>	
<b>Is a kit used</b>	yes
<b>Manufacturer name</b>	QIAGEN
<b>Specify the kit used</b>	DNeasy Plant Mini Kit
<b>Kit used following the manufacturer's instructions?</b>	
<b>Other information</b>	
<b>Other details on the test</b>	Commercial kit (DNeasy Plant Mini kit Qiagen) from leaf midribs or phloem tissue, previously powdered with liquid nitrogen. An alternative protocol has

	been used in the case of not availability of liquid nitrogen for the initial powdering of plant material. (Pasquini et al., 2013)
<b>Method: Molecular Conventional PCR</b>	
<b>Reference of the test description</b>	
<b>As or adapted from an EPPO diagnostic protocol</b>	yes
<b>EPPO Diagnostic Protocol name</b>	PM 7/062 Candidatus Phytoplasma mali (version 2)
<b>Name of the test</b>	AP group-specific nested PCR (adapted from Deng & Hiruki, 1991; Schneider et al., 1995; Lorenz et al., 1995)
<b>Other information</b>	
<b>Other details on the test</b>	Direct universal PCR with primers P1(Deng and Hiruki, 1991)/16S-Sr (Lee et al., 2004), followed by a nested 16SrX group specific with primers f01/rO1 (Lorenz et al., 1995)
<b>Are the performance characteristics included in the EPPO diagnostic protocol?</b>	<b>no</b>
<b>Performance Criteria :</b>	
<b>Organism 1.:</b>	<b>'Candidatus Phytoplasma mali'(PHYPMA)</b>
<b><u>Analytical sensitivity</u></b>	
<b>What is smallest amount of target that can be detected reliably?</b>	The analytical sensitivity was calculated analyzing three samples at seven dilution levels (1/1-1/1.000.000). The dilutions were in DNA from an healthy apple sample. Last dilution level with 100% positive results for all three samples: 1/1000
<b><u>Diagnostic sensitivity</u></b>	
<b>Proportion of infected/infested samples tested positive compared to results from the standard test, see appendix 2 of PM 7/98</b>	23 'target' samples: 20 apple samples infected by 'Ca. P. mali' coming from different Italian areas, 2 plum samples infected by 'Candidatus Phytoplasma prunorum' (ESFY phytoplasma) and one sample of pear infected by 'Candidatus Phytoplasma pyri' (PD phytoplasma) Diagnostic sensitivity: 83%
<b>Standard test(s)</b>	Other methodologies included in the ringtest: - Direct universal PCR with primers fAT/rAS (Smart et al., 1996) - SYBR Green real time PCR (Galetto et al., 2005)
<b><u>Analytical specificity - inclusivity</u></b>	
<b>Number of strains/populations of target organisms tested</b>	23 'target' samples: 20 apple samples infected by 'Ca. P. mali' coming from different Italian areas, 2 plum samples infected by 'Candidatus Phytoplasma prunorum' (ESFY phytoplasma) and one sample of pear infected by 'Candidatus Phytoplasma pyri' (PD phytoplasma)
<b>Specificity value</b>	Analytical specificity: 100%
<b><u>Analytical specificity - exclusivity</u></b>	
<b>Number of non-target organisms tested</b>	Two 'non target' samples were included:

	phytobacteria commonly spread on pome fruits: - Extracted DNA from a pear infected by Pseudomonas syringae pv. syringae - Extracted DNA from an apple infected by Erwinia amylovora
<b>Specificity value</b>	Not occurred
<b><u>Diagnostic Specificity</u></b>	
<b>Proportion of uninfected/uninfested samples (true negatives) testing negative compared to results from a standard test</b>	Five samples of apple tree uninfected (certified material) Diagnostic specificity: 100%
<b>Specify the test(s)</b>	Other methodologies included in the ringtest: - Direct universal PCR with primers fAT/rAS (Smart et al., 1996) - SYBR Green real time PCR (Galletto et al., 2005)
<b><u>Reproducibility</u></b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	The reproducibility was calculated analyzing in six laboratories all samples included in diagnostic specificity and sensitivity tests. Reproducibility: 89.1%
<b><u>Repeatability</u></b>	
<b>Provide the calculated % of agreement for a given level of the pest (see PM 7/98)</b>	The repeatability was calculated in three laboratories analyzing three samples at seven dilution levels (1/1-1/1.000.000). The dilutions were in DNA from an healthy apple sample. Repeatability: 100%
<b><u>Test performance study</u></b>	
<b>Test performance study?</b>	yes
<b>Brief details of the test performance study and its output. It available, link to published article/report</b>	A ringtest was organized with the official phytosanitary Italian laboratories within a Project financed by the Italian Ministry of Agriculture (ARNADIA) for the definition of 'Italian reference protocols'.

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